

Atherosclerosis and mitochondrial dysfunction – possible links

Magdalena Stachura¹, Stefan Grzegorz Pierzynowski²

¹ Cardiology Department with Cardiology Intensive Care Unit, Ministry of the Interior and Administration Hospital, Lublin, Poland

² Department of Cell and Organism Biology, Lund University, Lund, Sweden; Department of Medical Biology, Institute of Agricultural Medicine, Lublin, Poland

Abstract: Atherosclerosis is one of the most widespread diseases and remains a leading cause of death in developed countries. Traditional risk factors, such as obesity, cigarettes smoking, diabetes, and arterial hypertension, are insufficient for predicting which patients are in the group at highest risk for developing atherosclerosis. Knowledge about low-intensity inflammatory processes in the blood vessels, links between traditional risk factors and the forming of atherosclerotic plaques forming is constantly being extended, resulting in new strategies for treatment. Oxidative stress is described as an imbalance between reactive oxygen species (ROS) production and their elimination, in favour the former, and is believed to be characteristic of diabetes, hypertension and atherosclerosis. Increased level of ROS is considered an important cause of endothelial dysfunction occurring in the above-mentioned diseases by triggering the mitochondrial dysfunction. There is also evidence suggesting a possible link between mitochondrial dysfunction and cardiovascular diseases, thus indicating new directions for the examination of mechanisms of atherosclerosis.

Key words: atherosclerosis, oxidative stress, mitochondrial dysfunction, reactive oxygen species (ROS), reactive nitrogen species (RNS), endothelium dysfunction, 2-oxyglutarate

INTRODUCTION

Despite many preventive actions, atherosclerosis is still one of the most widespread diseases and remains a leading cause of death in developed countries. Knowledge about low-intensity inflammatory processes in the blood vessels, links between traditional risk factors, and the formation of atherosclerotic plaques forming is constantly being extended, resulting in strategies for new treatment. However, there are still many people who suffer from atherosclerosis complications, such as myocardial infarction, heart failure, brain stroke, peripheral arterial disease or renal failure – diseases which may significantly reduce physical ability and lower the quality of life.

Endothelium. The endothelium is the internal layer of blood vessels, and scientists formerly believed that it acted only as a passive barrier to blood elements. However, other endothelium functions, such as the regulation of vascular tone, supporting anticoagulative action and paracrine activity have been discovered recently. Endothelial permeability plays an important role in the maintaining of albumin gradients, necessary for fluid balance in the tissues, and is enhanced by angiotensin, interleukin 1 β or nitric oxide (NO). Vascular tone is regulated by endothelial secretion of vasodilator substances, such as NO and prostacyclines, and vasoconstrictor factors, e.g. angiotensin, endothelin-1, thromboxane A₂, and superoxide anion.

The endothelium prevents the formation of thrombus by the synthesis and secretion of anticoagulative substances

(antithrombin III, protein S, tissular factor inhibitor), platelets antiadherents (NO and prostacyclin), and fibrinolytic factors (plasminogen tissular factor), which is an advantage over the synthesis and secretion of procoagulative substances (von Willenbrand factor, fibronectin and thrombospondine). Another important role of endothelium is the synthesis and secretion of chemokines, activating the platelet factor, interleukin 8 (IL-8), and the chemotactic protein for monocytes (MCP-1), which enable lymphocytes and monocytes to migrate into the inner layer of arteries. Furthermore, vascular adhesion molecules, for example, vascular cellular adhesion molecule type 1 (VCAM-1), and intercellular adhesion molecule type 1 (ICAM-1), present on the surface of endothelial cells, also facilitate the transposition of leucocytes into the intima. In the presence of atherosclerosis risk factors, including cigarettes smoking, obesity, dyslipidemia, hypertension, and diabetes, the balance between different endothelium actions is disrupted. Such a status is called endothelium dysfunction and comprises the decrease of vasodilating and anticoagulative endothelial actions and enhanced proinflammatory molecules synthesis [1, 2, 3].

Oxidative stress. Oxidative stress is described as an imbalance between reactive oxygen species (ROS) production and their elimination, in favour the former, and is characteristic of diabetes, hypertension and atherosclerosis. An increased level of ROS is considered an important cause of endothelial dysfunction occurring in these diseases [2]. ROS generated in blood vessels include superoxide (O₂⁻), hydrogen peroxide (H₂O₂), hypochlorous acid, hydroxyl radicals ([•]OH) and singlet oxygen (¹O₂), with superoxide being the most importance [3]. ROS can directly inactivate endothelial-derived NO, cause protein dysfunction and cell signaling disturbances, which initiate and escalate endothelial dysfunction [2]. Many

Corresponding author: Dr. Magdalena Stachura, Cardiology Department with Cardiology Intensive Care Unit, Ministry of the Interior and Administration Hospital, Grenadierów 3, 20-331 Lublin, Poland.
E-mail: magdas10@onet.eu

Received: 16 December 2009; accepted: 31 December 2009

enzymes, including NADPH oxidases, xanthine oxidase (XO), myeloperoxidase (MPO), and nitric oxide synthases (NOS), are sources of ROS in vessels. However, the leakage of electrons from mitochondrial electron-transport chain is believed to be a major source of ROS [3]. Elevated inner mitochondrial membrane potential, calcium ions, and NO are expected to regulate electron-transport chain ROS production [4]. Complex III of the electron-transport chain is also believed to be a major source of ROS. On the other hand, the Krebs cycle enzymes, especially 2-oxoglutarate dehydrogenase and pyruvate dehydrogenase, are also expected to generate ROS in the condition of increased NADH/NAD⁺ ratio [3].

Reactive nitrogen species (RNS) generated in the vascular system, such as peroxynitrite (ONOO⁻), a product of the reaction between superoxide and NO, are also considered as a cause of cell damage and death. Both ROS and RNS may cause oxidative modifications of low-density lipoproteins (LDL), thus enhancing the formation of atherosclerotic plaques [1, 2, 3].

Antioxidants. There are several mechanisms of antioxidant actions. Intracellular antioxidants include superoxide dismutase (SOD) enzymes, which are metalloenzymes enabling the conversion of O₂^{•-} to H₂O₂. One of the SOD isoforms contains copper and zinc, and is located in the cytoplasm, another isoform is mitochondrial and contains manganese, while another SOD isoform is located in extracellular matrix, has been described. Reduced glutathione (GSH) peroxidase and catalase enable the conversion of H₂O₂ to H₂O in a condition that reduced glutathione (GSH) is available. GSH also acts as a direct antioxidant due to hydrogen ions donation [3].

According to present knowledge, ROS and RNS play an important regulatory role, which consists in the activation of mechanisms that control cell differentiation and the apoptosis processes [2, 3]. These processes are described as redox signaling and depend on posttranslational proteins modifications. It is known that ROS and RNS may also influence transcriptional factors activity. Among the transcriptional factors which remain under redox control are: nuclear factor- κ B (NF- κ B); hypoxia-induced factor 1 (HIF-1); nuclear factor (erythroid-derived 2)-like 2 (nrf2); and activator protein 1 (AP-1). In endothelial cells H₂O₂ improves the activity of NF κ B [5], which results in enhanced inflammatory response [2].

Mitochondrial damage. In conditions of oxidative stress, antioxidant defence systems are not sufficient to neutralize excessive amounts of ROS and RNS, which leads to protein, lipid, and DNA damage. Mitochondria are the most exposed to ROS and RNS organelles, and are also expected to be the most susceptible to damages caused by ROS and RNS [3, 6]. ROS and RNS are produced in mitochondria close to mitochondrial DNA, which is due to the lack of histones less prevented from oxidative damage than nuclear DNA. Also mitochondrial polymerases are more susceptible to caused by ROS modifications [3]. ROS may also induce cardiolipin oxidation due to its high content of unsaturated fatty acids and its close location to mitochondrial electron-transport chain [7], and due also to its decreasing complex I activity, leading to cytochrome c release [8].

According to recent data, mitochondrial dysfunction triggers cell necrosis and apoptosis pathways, thus playing a significant role in the development of diseases such as obesity, diabetes, heart failure, stroke, neurodegenerative diseases and cancer [3]. There is also evidence suggesting a possible link between

mitochondrial dysfunction and cardiovascular diseases. In apolipoprotein E (apoE) knockout mice mitochondrial DNA oxidative damage was positively correlated with the extension of atherosclerotic plaques [9]. According to clinical data, patients with atherosclerosis, such as people with elevated cholesterol blood levels and smoking habit, revealed more disturbances in mitochondrial DNA in samples of their heart and arterial tissues than healthy controls [9, 10].

Oxidized lipids. Oxidized lipids may initiate cellular responses through either receptor-mediated or post-translational proteins modifications. It has been proved that low levels of oxidized lipids have a cytoprotective influence on cells, but high concentrations of oxidized lipids lead to cell apoptosis. Oxidized lipids are also expected to impair mitochondrial function [1, 3]. Endothelial cells incubation with oxidized LDL led to the induction of a mitochondrial complex I activity expected to depend on oxidative stress induction [11]. Recent data suggest that oxidized LDL may influence even more mitochondrial respiratory chain enzymes activity. Cultured porcine aortic endothelial cells incubation with oxidized LDL resulted in the decrease of NADH-ubiquinone dehydrogenase (complex I of mitochondrial respiratory chain), succinate cytochrome c oxidase (complex II/III), ubiquinone cytochrome c reductase (complex III), cytochrome c oxidase (complex IV), such as NAD⁺/NADH ratio [12].

Another study revealed the induction the transcription and expression of mitochondrial SOD in human macrophages incubated with oxidized LDL [13]. It has also been proved that human macrophage incubation with oxidized LDL leads to enhanced production of mitochondrial ROS, and lowers mitochondrial membrane potential [14]. In another study, treatment of rat smooth muscle cells with oxidized LDL lowered intracellular ATP levels, such as mitochondrial oxidative phosphorylation subunits mRNA expression, impaired mitochondrial electron-transport chain capacity, insulin-mediated phosphorylation of Akt and AMP-activated protein kinase (AMPK). Mitochondrial dysfunction stimulated the migration abilities of these cells, probably through the inactivation of Akt, which may play a crucial role in the formation of atherosclerotic lesions [15]. However, increased production of ROS leads to vascular smooth muscle cells and macrophages apoptosis, thus resulting in the destabilisation of atherosclerotic plaques [16]. The destabilization of atherosclerotic lesions enables thrombus forming, which plays crucial role in the development of acute heart ischemia, which leads to myocardial infarction. Ischemia conditions trigger anaerobic metabolic pathways in heart tissue, which include the utilization of anaerobic glycolysis and fatty acids, enhanced glucose uptake, and decrease of heart muscle contractility.

In the case of persisting ischemia, the compensative mechanisms become insufficient and heart tissue develops significant ATP deficit; consequently resulting in increased calcium ions flow into the cell cytoplasm, xanthine oxidase activation, and increased ROS generation leading to mitochondrial dysfunction [17, 18]. On the other hand, also reperfusion processes are involved with increased free radicals production, due to the leakage of electrons from mitochondrial electron transport chain and xanthine oxidase activation [17].

Mitochondrial dysfunction leads to increased ROS and RNS generation, which results in an enhanced intravascular

inflammatory response, impaired endothelial NO synthesis, the dysfunction of the endothelium, and the initiation and progression of atherosclerosis. However, inflammatory cytokines, such as tumour necrosis factor- α (TNF- α) enhance mitochondrial dysfunction through the increase of NAD(P)H oxidase activity and ROS production [19]. Data suggest that also NO may play a regulative role in mitochondrial oxidative stress protection through an influence on peroxisome proliferator-activated

Mice with switched-off endothelial NO synthase activity revealed reduced levels of PGC-1 α and reduced expression of genes involved in mitochondrial oxidative stress protection mechanisms, such as catalase, manganese SOD, peroxiredoxin III, peroxiredoxin V, thioredoxin 2 and thioredoxin reductase 2 [20]. Data also suggest that nitrite therapy after cardiac arrest protects against increased ROS generation during the reperfusion processes. In this experiment, nitrite-treated mice revealed a reversible inhibition of complex I of electron transport chain system resulting in decreased levels of reperfusion ROS production, but without any reduction in electron transport chain efficiency [21].

The Krebs cycle. According to recent data, the Krebs cycle inhibition seems to be an early marker of endothelium dysfunction. Chronic inhibition of NO synthesis in mice results not only, as expected, in elevated levels of endothelial dysfunction markers such as soluble ICAM-1, VCAM-1 and matrix metalloproteinase 9 levels, but also in the selective inhibition of Krebs cycle enzymes: aconitase-2 and enoyl-CoA-hydratase-1, accompanied by reduced mitochondrial mass. These findings suggest that endothelial dysfunction results in Krebs cycle inhibition and enhanced glycolytic pathway of pyruvate usage, similarly to the hypoxia conditions [22]. 2-oxoglutarate dehydrogenase is another Krebs cycle enzyme which catalyses the conversion of 2-oxoglutarate to succinyl-CoA, thus producing NADH and providing electrons for electron-chain transport system, is expected to be also influenced by oxidative stress. As a result of increased ROS production, 2-oxoglutarate dehydrogenase inhibiting significantly lowers the supply of NADH to respiratory chain. On the other hand 2-oxoglutarate dehydrogenase also generates ROS by itself, when NADH/NAD⁺ ratio is elevated [23].

2-oxoglutarate. 2-oxoglutarate (2-OG) is the Krebs cycle intermediate which is converted in the mitochondria into succinyl-CoA by 2-oxoglutarate dehydrogenase. Moreover, 2-OG is also contained in the cytoplasm and blood plasma where it is expected to act as a free ammonia scavenger, such as participating in proline synthesis in the intestine, and proline conversion to hydroxyproline [24, 25]. The origin of cytoplasmic and plasma 2-OG needs further explanation, but it has been proved that plasma 2-OG levels decrease together with the increase in the age of the patients [26].

According to recent data, oral 2-OG administration improved arterial wall elasticity and led to the increase of total collagen content in the walls of arteries in elderly mice [27]. Another study revealed that 2-OG treatment protects from oxidative stress cataract formation induced in rats. This suggests that 2-OG may act as ROS scavenger due to its alpha-keto-carboxylate group [28]. Another study proved the positive influences of 2OG on lipid peroxidation and antioxidant status in rats treated with ammonium acetate. In the study, rats

treated with 2-OG were protected from developing metabolic disorders, such as the increase in free fatty acids, triglycerides, phospholipids, cholesterol, serum transaminases, and thiobarbituric acid reactive substances plasma levels, caused by ammonium acetate administration [29]. Data also suggest, that oral 2OG administration has beneficial influence on blood lipid levels. The study revealed that treatment with 2OG led to the decrease of total cholesterol, LDL and triglycerides in plasma, and the increase of HDL blood level in rats with experimentally-induced hyperlipidemia [30]. Recent data suggest that postnatal 2-OG administration has a protective influence on lipid metabolism in piglets prenatally exposed to dexamethasone. In the study, treatment with 2-OG resulted in a 40% reduction of total cholesterol plasma level, compared with the control group [31].

To summarise, atherosclerosis is one of the most widespread diseases in developed countries. Endothelium dysfunction is believed to play an essential role in the initiation and progression of formation of atherosclerotic lesions, and that oxidative stress is one of the most important risk factors that evokes and enhances endothelium function disorders, although knowledge about the links between traditional risk factors and the development of atherosclerosis continually comes to the fore. However, as long as it remains unpredictable which patients are in the group with the highest acute risk for coronary syndrome, and require the most aggressive treatment, and despite advanced treatment methods, it is still not possible to prevent some patients from developing atherosclerosis complications, further investigations are required.

REFERENCES

1. Victor VM, Rocha M, Solá E, Bañuls C, Garcia-Malpartida K, Hernández-Mijares A: Oxidative stress, endothelial dysfunction and atherosclerosis. *Curr Pharm Des* 2009, **15**(26), 2988-3002.
2. Thomas SR, Witting PK, Drummond GR: Redox control of endothelial function and dysfunction: molecular mechanisms and therapeutic opportunities. *Antioxid Redox Signal* 2008, **10**(10), 1713-1765.
3. Victor VM, Rocha M: Targeting antioxidants to mitochondria: a potential new therapeutic strategy for cardiovascular diseases. *Curr Pharm Des* 2007, **13**(8), 845-863.
4. Moncada S, Erusalimsky JD: Does nitric oxide modulate mitochondrial energy generation and apoptosis? *Nat Rev Mol Cell Biol* 2002, **3**(3), 214-220.
5. Csiszar A, Smith K E, Koller A, Kaley G, Edwards JG, Ungvari Z: Regulation of bone morphogenetic protein-2 expression in endothelial cells: role of nuclear factor-kappaB activation by tumor necrosis factor- α , H₂O₂, and high intravascular pressure. *Circulation* 2005, **111**(18), 2364-2372.
6. Victor VM, Rocha M: Targeting antioxidants to mitochondria: a potential new therapeutic strategy for cardiovascular diseases. *Curr Pharm Des* 2007, **13**(8), 845-863.
7. Paradies G, Petrosillo G, Pistolesi M, Di Venosa N, Federici A, Ruggiero FM: Decrease in mitochondrial complex I activity in ischemic/reperfused rat heart: involvement of reactive oxygen species and cardiolipin. *Circ Res* 2004, **94**(1), 53-59.
8. Nomura K, Imai H, Koumura T, Kobayashi T, Nakagawa Y: Mitochondrial phospholipid hydroperoxide glutathione peroxidase inhibits the release of cytochrome c from mitochondria by suppressing the peroxidation of cardiolipin in hypoglycaemia-induced apoptosis. *Biochem J* 2000, **351**(Pt 1), 183-193.
9. Ballinger SW, Patterson C, Knight-Lozano CA, Burow DL, Conklin CA, Hu Z, Reuf J, Horaist C, Lebovitz R, Hunter GC, McIntyre K, Runge MS: Mitochondrial integrity and function in atherogenesis. *Circulation* 2002, **106**(5), 544-549.
10. Knight-Lozano CA, Young CG, Burow DL, Hu ZY, Uyeminami D, Pinkerton KE, Ischiropoulos H, Ballinger SW: Cigarette smoke exposure and hypercholesterolemia increase mitochondrial damage in cardiovascular tissues. *Circulation* 2002, **105**(7), 849-854.

11. Ceaser EK, Ramachandran A, Levenon AL, Darley-Usmar VM: Oxidized low-density lipoprotein and 15-deoxy-delta 12,14-PGJ2 increase mitochondrial complex I activity in endothelial cells. *Am J Physiol Heart Circ Physiol* 2003, **285**(6), H2298-2308.
12. Roy Chowdhury SK, Sangle GV, Xie X, Stelmack GL, Halayko AJ, Shen GX: Effects of extensively oxidized low-density lipoprotein on mitochondrial function and reactive oxygen species in porcine aortic endothelial cells. *Am J Physiol Endocrinol Metab* 2009 Oct 20. [Epub ahead of print]
13. Kinscherf R, Deigner HP, Usinger C, Pill J, Wagner M, Kamencic H, Hou D, Chen M, Schmiedt W, Schrader M, Kovacs G, Kato K, Metz J: Induction of mitochondrial manganese superoxide dismutase in macrophages by oxidized LDL: its relevance in atherosclerosis of humans and heritable hyperlipidemic rabbits. *FASEB J* 1997, **11**(14), 1317-1328.
14. Asmis R, Begley JG: Oxidized LDL promotes peroxide-mediated mitochondrial dysfunction and cell death in human macrophages: a caspase-3-independent pathway. *Circ Res* 2003, **92**(1), e20-29.
15. Ahn SY, Choi YS, Koo HJ, Jeong JH, Park WH, Kim M, Piao Y, Pak YK: Mitochondrial dysfunction enhances the migration of vascular smooth muscle cells via suppression of Akt phosphorylation. *Biochim Biophys Acta* 2009 Sep 23. [Epub ahead of print]
16. Madamanchi NR, Runge MS: Mitochondrial dysfunction in atherosclerosis. *Circ Res* 2007, **100**(4), 460-473.
17. Misra MK, Sarwat M, Bhakuni P, Tuteja R, Tuteja N: Oxidative stress and ischemic myocardial syndromes. *Med Sci Monit* 2009, **15**(10), RA209-219.
18. Chambers DE, Parks DA, Patterson G, Roy R, McCord JM, Yoshida S, Parmley LF, Downey JM: Xanthine oxidase as a source of free radical damage in myocardial ischemia. *J Mol Cell Cardiol* 1985, **17**(2), 145-112.
19. Sprague AH, Khalil RA: Inflammatory cytokines in vascular dysfunction and vascular disease. *Biochem Pharmacol* 2009, **78**(6), 539-552.
20. Borniquel S, Valle I, Cadenas S, Lamas S, Monsalve M: Nitric oxide regulates mitochondrial oxidative stress protection via the transcriptional coactivator PGC-1alpha. *FASEB J* 2006, **20**(11), 1889-1891.
21. Dezfulian C, Shiva S, Alekseyenko A, Pendyal A, Beiser DG, Munasinghe JP, Anderson SA, Chesley CF, Vanden Hoek TL, Gladwin MT: Nitrite therapy after cardiac arrest reduces reactive oxygen species generation, improves cardiac and neurological function, and enhances survival via reversible inhibition of mitochondrial complex I. *Circulation* 2009, **120**(10), 897-905.
22. Addabbo F, Ratliff B, Park HC, Kuo MC, Ungvari Z, Csiszar A, Krasnikov B, Sodhi K., Zhang F, Nasjletti A, Goligorsky MS: The Krebs cycle and mitochondrial mass are early victims of endothelial dysfunction: proteomic approach. *Am J Pathol* 2009, **174**(1), 34-43.
23. Tretter L, Adam-Vizi V: Alpha-ketoglutarate dehydrogenase: a target and generator of oxidative stress. *Philos Trans R Soc Lond B Biol Sci* 2005, **360**(1464), 2335-2345.
24. Filip RS, Pierzynowski SG: The role of glutamine and alpha-ketoglutarate in gut metabolism and the potential application in medicine and nutrition. *J Pre-Clin Clin Res* 2007, **1**, 1-7.
25. Filip RS, Pierzynowski SG, Lindegard B, Wernerman J, Haratym-Maj A, Podgurniak M.: Alpha-ketoglutarate decreases serum levels of C-terminal cross-linking telopeptide of type I collagen (CTX) in postmenopausal women with osteopenia: six-month study. *Int J Vitam Nutr Res* 2007, **77**(2), 89-97.
26. Harrison AP, Pierzynowski SG: Biological effects of 2-oxoglutarate with particular emphasis on the regulation of protein, mineral and lipid absorption/metabolism, muscle performance, kidney function, bone formation and cancerogenesis, all viewed from a healthy ageing perspective state of the art – review article. *J Physiol Pharmacol* 2008, **59** (Suppl 1), 91-106.
27. Harrison A, Bruggemann D, Bartels EM, Andrea K, Pierzynowski S: Helathy ageing: the beneficial effect of dietary supplementation with alpha-ketoglutarate on arterial elasticity in elderly mice. *Aging Cell* 2009, article in press.
28. Varma SD, Hegde KR: Effect of alpha-ketoglutarate against selenite cataract formation. *Exp Eye Res* 2004, **79**(6), 913-918.
29. Velvizhi S, Dakshayani KB, Subramanian P: Protective influences of alpha-ketoglutarate on lipid peroxidation and antioxidant status in ammonium acetate treated rats. *Indian J Exp Biol* 2002, **40**(10), 1183-1186.
30. Radzki RP, Bieńko M, Pierzynowski SG: Effect of dietary alpha-ketoglutarate on blood lipid profile during hypercholesterolaemia in rats. *Scand J Clin Lab Invest* 2009, **69**(2), 175-180.
31. Śliwa E, Dobrowolski P, Tataro MR, Pierzynowski SG: Alpha-ketoglutarate partially protects newborns from metabolic changes evoked by chronic maternal exposure to glucocorticoids. *J Pre-Clin Clin Res* 2008, **1**(1), 55-59.